



Nanostructured Metal Organic Framework Modified Glassy Carbon Electrode as a High Efficient Non-Enzymatic Amperometric Sensor for Electrochemical Detection of H₂O₂

Maryam Naseri¹, Lida Fotouhi^{1*}, and Ali Ehsani^{2*}

¹Department of Chemistry, Alzahra University, Tehran, Iran

²Department of Chemistry, Faculty of Science, University of Qom, Qom, Iran

ABSTRACT

Metal-organic frameworks have recently been considered very promising modifiers in electrochemical analysis due to their unique characteristics among which tunable pore sizes, crystalline ordered structures, large surface areas and chemical tenability are worth noting. In the present research, Cu(btec)_{0.5}DMF was electrodeposited on the surface of glassy carbon electrode at room temperature under cathodic potential and was initially used as the active materials for the detection of H₂O₂. The cyclic voltammogram of Cu(btec)_{0.5}DMF modified GC electrode shows distinct redox peaks potentials at +0.002 and +0.212 V in 0.1 M phosphate buffer solution (pH 6.5) corresponding to Cu^(II)/Cu^(I) in Cu(btec)_{0.5}DMF. Acting as the electrode materials of a non-enzymatic H₂O₂ biosensor, the Cu(btec)_{0.5}DMF brings about a promising electrocatalytic performance. The high electrocatalytic activity of the Cu(btec)_{0.5}DMF modified GC electrode is demonstrated by the amperometric response towards H₂O₂ reduction with a wide linear range from 5 μM to 8000 μM, a low detection limit of 0.865 μM, good stability and high selectivity at an applied potential of -0.2 V, which was higher than some H₂O₂ biosensors.

Keywords : MOF, Electrochemical, Biosensor, Amperometry

Received : 7 July 2017, Accepted : 22 January 2018

1. Introduction

The precise and quick detection of H₂O₂ is of practical importance in clinical diagnostics areas, food industry and biotechnology. One can name various techniques for the determination of H₂O₂, including titrimetry, spectrophotometry, fluorescence, chemiluminescence and electrochemistry [1-3]. Among the techniques mentioned above, the electrochemical one has been extensively utilized due to its simplicity, low-cost, high sensitivity and selectivity. During the past decades, most of the modified electrodes for determination of H₂O₂ were based on enzyme because of their high sensitivity and excellent selectivity. However, these expensive enzyme-based elec-

trochemical biosensors cannot be utilized in extreme weather and the detecting accuracy is greatly affected by factors like temperature, humidity, pH and toxic chemicals. By the same token, non-enzymatic electrochemical biosensors are good choices for H₂O₂ detection because of low cost, free of pH, high sensitivity, high stability and fast response time.

A new type of crystalline porous materials, metal-organic frameworks (MOFs) have attracted a great bulk of attention because of their unique properties such as high surface areas and high pore volumes in uniformly sized. The electrodeposition method has proven itself as the most promising technique among different methods of preparing thin films of MOFs owing to its simple procedure, easy handling, high yield efficiency, high purity of products and very short reaction time. In particular, MOFs have been considered useful modifiers in electrochemical analysis, providing novel electrochemical sensors. In this

*E-mail address: lfotouhi@alzahra.ac.ir (L. Fotouhi),
ehsani46847@yahoo.com (A. Ehsani)

DOI: <https://doi.org/10.5229/JECST.2018.9.1.28>

aspect, Wang's team reported an electrochemical sensor based on a carbon paste electrode (CPE), modified with $\text{ZnO}_4(\text{BDC})_3(\text{MOF-5})$ (BDC = 1,4-benzene dicarboxylate) for sensitive determination of lead. They applied a bulk modification of CPE instead of plating a film on the electrode surface because Zn-based MOFs are susceptible to water and lose their large surface area when exposed to water [4]. In a similar study, they prepared amino functionalized metalorganic frameworks ($\text{NH}_2\text{-Cu}_3(\text{BTC})_2$) (BTC = 1,3,5-benzene tricarboxylate) and introduced a novel electrode modifier to determine trace levels of lead [5]. Recently, a non-enzymatic H_2O_2 biosensor based on Ni(II)-based metal-organic framework (Ni(II)-MOFs) anchored on carbon nanotubes (CNTs) was made by Bao et al. [6].

Owing to the advantages of the MOFs, $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ (Cu-MOF ; btec = 1,2,4,5-benzene tetracarboxylate) (Fig. 1) was electrodeposited on the surface of glassy carbon electrode (GCE) in room temperature under cathodic potential and was initially employed as the active material for the detection of H_2O_2 in this report. Electrosynthesized $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ structure was characterized by scanning electron microscopy (SEM), transmission electron microscopy (TEM) and fourier transform infrared spectroscopy (FT-IR). This synthetic protocol supplies high surface area, high purity, easy of synthesis procedure, low cost and easy operation. As far as we know, it is the first example in which $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ is used as a novel electrocatalyst for H_2O_2 electrooxidation. The $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ modified GC electrode gives a wide linear range of 5-8000 μM , a good detection limit and high selectivity to the target analysts. As far as we know, such catalytic activities of Cu-MOFs have been rarely reported. Therefore, this Cu-MOF is very likely to be applied successfully in H_2O_2 non-enzymatic electrochemical sensors.

2. Experimental

2.1. Reagents and Materials

All reagents were bought from the Merck and Aldrich chemical companies and were used without any additional purification. Double distilled water was used during the experiments.

2.2. Apparatus

Electrochemical measurements were done by

Zahner Ennium and a conventional three-electrode system was employed during this experiment. A saturated Ag/AgCl electrode, a Pt wire and a glassy carbon electrode (2 mm diameter) were used to serve as the reference, counter and working electrodes, respectively. The frequency range of 100 kHz to 100 mHz and modulation amplitude of 5 mV were employed for impedance studies. The capacitance values were normalized by fitting on Z-view program. Tescan scanning electron microscope, energy dispersive X-ray spectrometry (EDX), and Zeiss EM900 transmission electron microscope have been used to obtain the morphologic information of sample. Fourier transform infrared spectra were recorded using a Bruker Tensor 27 FT-IR spectrometer with KBr pellets. Outgassing and determination of adsorption isotherms were done by a BEL Japan, Inc. surface area analyzer. Samples were outgassed at 120°C overnight under vacuum and N_2 adsorption isotherms were measured at 77 K.

2.3. Electrodeposition of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$

The $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ was electrochemically synthesized based on the previous report [7]. In short, the optimal values of organic linker (0.1 M H_4btec), copper nitrate (0.02 M), sodium nitrate (0.1 M) and supporting electrolyte (0.1 M tetrabutylammonium tetrafluoroborat (TBTA)) were dissolved in DMF:water (100:1 V/V) solution. A saturated Ag/AgCl electrode, a Pt wire and a glassy carbon electrode (2 mm diameter) were selected to be the reference, counter and working electrodes. Before the experiment, the GCE was cleaned by polishing with 0.05 μm alumina slurry on a polishing cloth to create a mirror finish. The electrode was then sonicated with absolute ethanol and double-distilled water for about 2.0 minutes, respectively, and finally, it was dried under ambient temperature. Electrolysis was carried out in an electrochemical cell under optimal constant voltage electrolysis (-1.7 V) for 10 minutes. Finally, the green precipitate of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ was deposited on the GCE. After that, a 5 μL of nafion aqueous solution (0.5 wt%) was coated on the modified electrode and was dried under ambient temperature.

3. Results and Discussion

3.1. Characterization of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$

The transmission electron microscopy and scan-

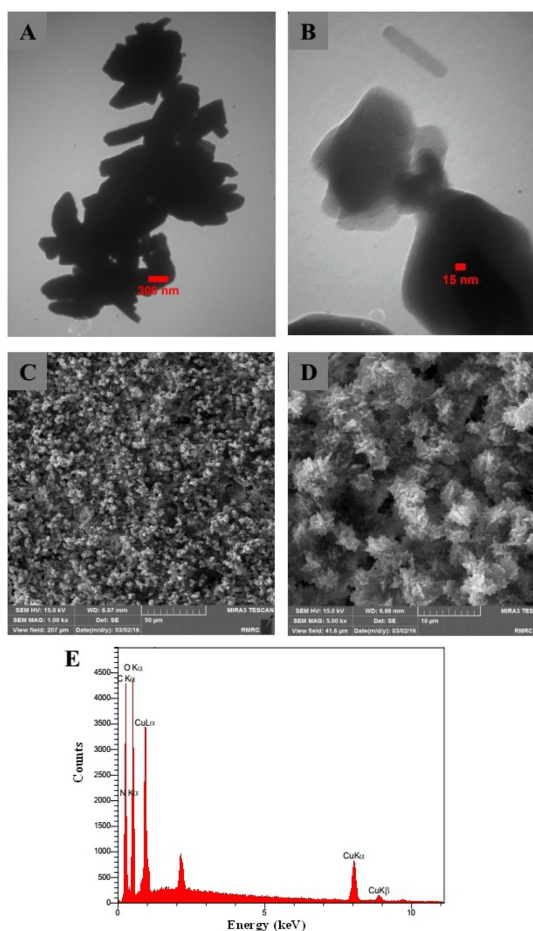


Fig. 1. TEM, SEM images and EDX spectrum of electro synthesized $\text{Cu}(\text{btec})_{0.5}\text{DMF}$.

ning electron microscopy characterize the microstructure and morphology of electro synthesized $\text{Cu}(\text{btec})_{0.5}\text{DMF}$. As shown in Fig. 1A, the $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ particles are rods particles in shape and are very similar in size. Fig. 1B shows a closer view of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ nano rods at the high-magnification TEM image. Moreover, it is interesting to see that SEM images of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ particles strongly resemble the natural flowers in Fig. 1C (shown in a large scale). As it can be seen in Fig. 1C, the morphology of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ to be uniform nanoflower arrays with monomodal particle size distribution of 40-50 nm on average. Fig. 1D indicates a high-magnification of an assembled 3D flower nanostructure particle. Moreover, the EDX analysis has shown that the $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ consisted of C, O, Cu and N elements. The N element in the spectrum should be from DMF, showing that $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ had been synthesized successfully.

The N_2 adsorption-desorption isotherm of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ is shown in Fig. 2A. The fractal dimension of solid samples can be determined based on the Frenkel-Halsey-Hill (FHH) method by utilizing the isotherm data of N_2 gas adsorption. The equation used in FHH method is:

$$\ln\left(\frac{V}{V_0}\right) = \text{constant} + A \left[\ln\left(\frac{P_0}{P}\right)\right] \quad (1)$$

where V and V_0 are the volume of adsorbed gas at pressure P and monolayer volume, respectively. P_0 and A , respectively, are the saturation pressure and a constant, related to the fractal dimension (D_f) of solid

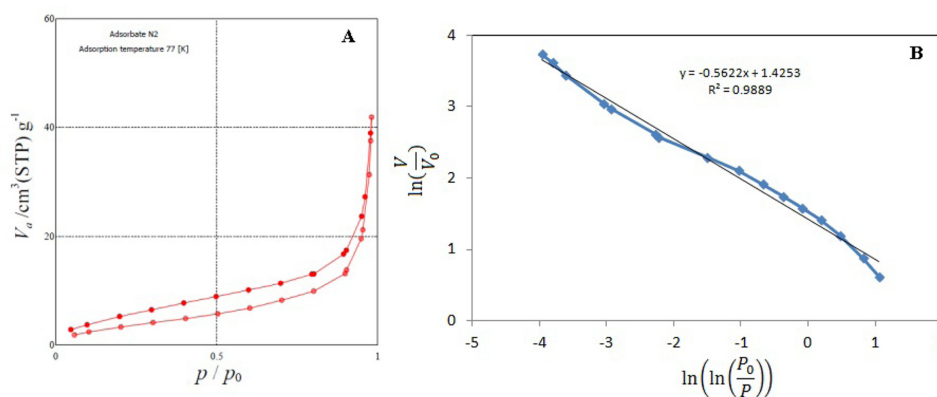


Fig. 2. (A) The N_2 adsorption-desorption isotherm of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ and (B) The fractal dimension plot of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$.

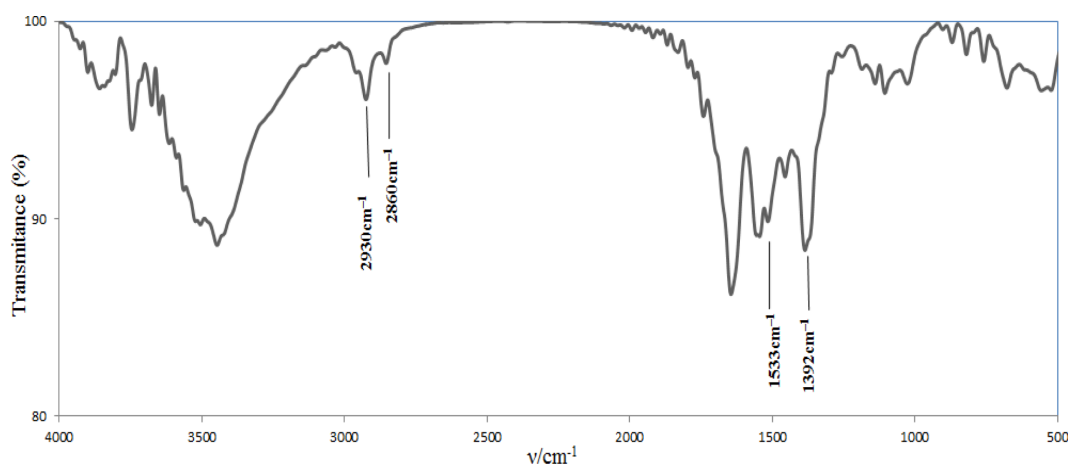


Fig. 3. Infrared spectra of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$.

sample ($D_f = A + 3$). As far as FHH method is concerned, the plot of $\ln(V)$ versus $\ln(\ln(P_0/P))$ gives a straight line with slope of A . The fractal dimension plot of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ is shown in Fig. 2B. In this regards, fractal dimension values of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ was calculated as 2.44.

Infrared spectra of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ is shown in Fig. 3. The $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ displays the absorption bands at 1533 cm^{-1} and 1392 cm^{-1} , which is related to the antisymmetric stretching vibrations and symmetric stretching vibrations of $-\text{COO}^-$, respectively. No absorption peaks are observed between 1730 cm^{-1} and 1690 cm^{-1} , indicating that all the carboxyl groups are deprotonated. The appearance of stretching vibrations of C-H bands (related to methyl group in DMF molecule) at 2930 cm^{-1} and 2860 cm^{-1} is a good indication of the presence of DMF in the structure of the electrosynthesized $\text{Cu}(\text{btec})_{0.5}\text{DMF}$.

3.2. Electrochemical Performance of $\text{Cu}(\text{btec})_{0.5}\text{DMF-GCE}$

For the electronic conductivity of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ to be analyzed, electrochemical impedance spectroscopy was used. The bare GCE Nyquist plots and modified GCE are displayed in Fig. 4. The semicircle portion of the plot corresponds to the charge transfer process, with the diameter of the semicircle equivalent to the charge transfer resistance (R_{ct}). After modification with $\text{Cu}(\text{btec})_{0.5}\text{DMF}$, the EIS semicircle diameter of $\text{Cu}(\text{btec})_{0.5}\text{DMF-GCE}$ is markedly reduced. The electron transfer rate constant (k_{et}) at the $\text{Cu}(\text{btec})_{0.5}\text{DMF-GCE}$ interface can be estimated

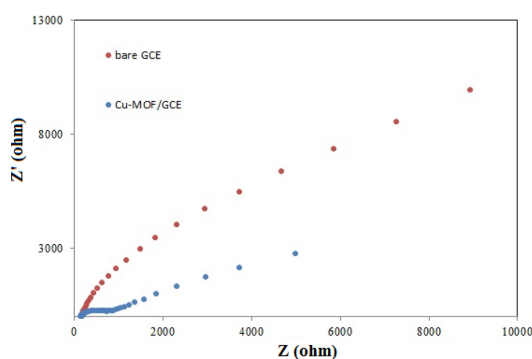


Fig. 4. The Nyquist plots of bare GCE and modified GCE in 0.1 M PBS.

by using the EIS data and the equation as follows [8]:

$$k_{et} = \frac{1}{2(R_{ct})(CPE)} \quad (2)$$

where CPE is constant phase element and R_{ct} is the charge transfer resistance. The values of CPE and R_{ct} were obtained by fitting on Z-view program as $1.944\text{ }\mu\text{Fs}^{-1}$ and $1148\text{ }\Omega$, respectively. Then the electron transfer rate constant, $k_{et} = 224$, was calculated based on Eq. 2. These results demonstrate that $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ can form good electron pathways between the electrode and electrolyte; thus, this can serve as an appropriate platform for electrocatalysis applications.

The electrochemical behavior of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$, which is electrodeposited on GC electrode, was studied by CV. Fig. 5A indicate the CV response of

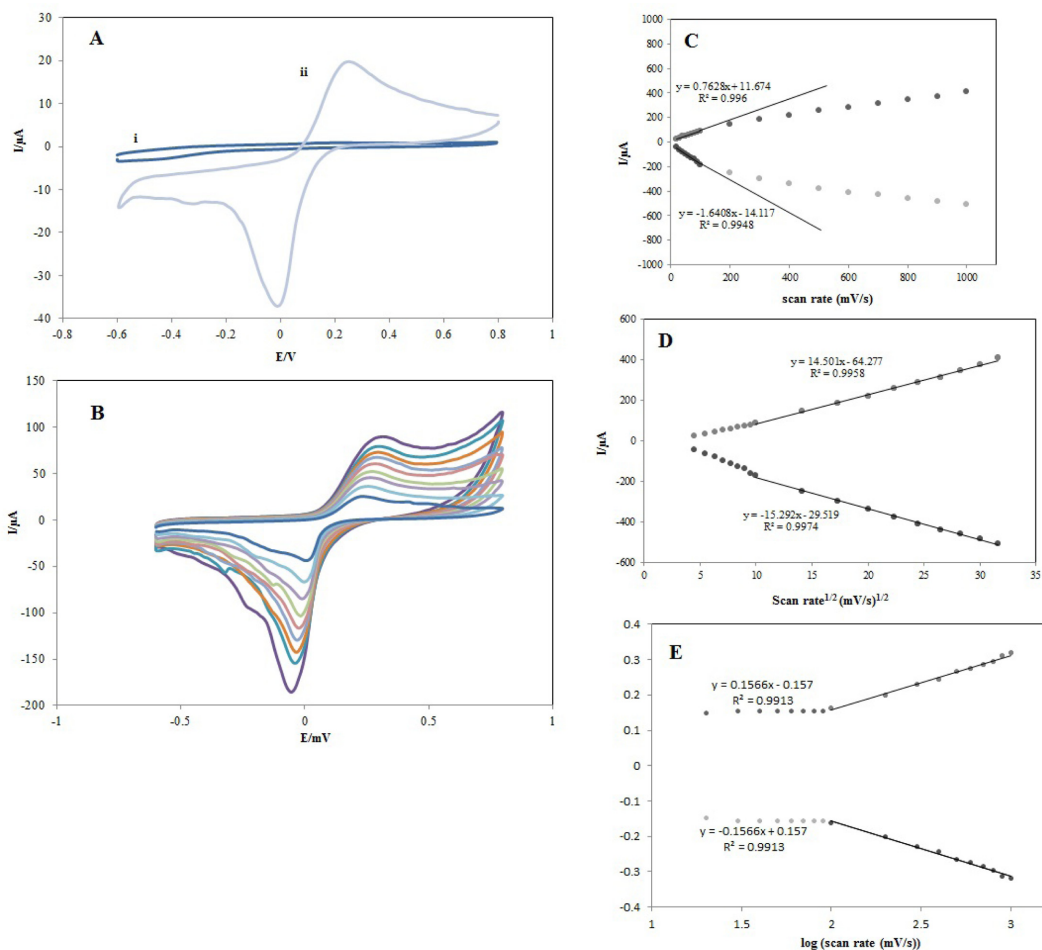


Fig. 5. (A) CV response of $\text{Cu}(\text{btec})_{0.5}\text{DMF}/\text{GCE}$ in 0.1 M phosphate buffer solution (PBS) at a scan rate of 30.0 mV s^{-1} at pH 6.5, (B) CVs of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ electrodeposited at different scan rates, (C) dependency of the peak currents of the redox system vs scan rate below 100 mV s^{-1} (D) dependency of the peak currents of the redox system vs square root of the scan rate and (E) shift values of the peak potentials vs to logarithm of the scan rates.

$\text{Cu}(\text{btec})_{0.5}\text{DMF}/\text{GCE}$ in 0.1 M phosphate buffer solution (PBS) at a scan rate of 30.0 mV s^{-1} at pH 6.5. As shown in the figure, the resulting cyclic voltammogram arrives at distinct redox peak potentials at $+0.002$ and $+0.212 \text{ V}$ which was presumably ascribed to the reversible oxidation and reduction of $\text{Cu}^{(II)}/\text{Cu}^{(I)}$ in $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ [9]. In order to study the kinetic parameters, the CVs of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ electrodeposited on a glassy carbon electrode at different scan rates in 0.1 M PBS (pH 6.5) were recorded (Fig. 5B). The redox system's peak currents are proportional to the scan rate for sweep rates below 100 mV s^{-1} (Fig. 5C) signaling a surface-

controlled process. In addition, the anodic peak currents were quite similar to the corresponding cathodic peak currents and the peak potential did not change with the increase in the scan rate. The plot of the peak current vs. scan rate, at higher sweep rates, deviates from linearity and, thus, the peak current becomes proportional to the square root of the scan rate (Fig 5D), as in redox diffusion-controlled process. Peak separations, at higher sweep rates ($>100 \text{ mV s}^{-1}$), begin to increase and the shift values of the peak potentials are proportional to the logarithm of the scan rate (Fig. 5E). Charge transfer coefficient (α) and the electron transfer rate constant (k_s), based on the Laviron the-

ory, can be arrived at by measuring the variation of peak potential by scan rate when $n\Delta E_p \geq 200$ mV [10]. The charge transfer coefficient was determined by using the following equation:

$$E_p = K - 2.3030 \frac{RT}{\alpha nF} \log v \quad (3)$$

The plot of ΔE_p vs. $\log(v)$ displays a slope about 156 mV and, then, α was calculated as 0.378 (for $n = 1$). The electron transfer rate constant can be obtained by introducing this value in the following equation:

$$\text{Log}k_s = \alpha \log(1-\alpha) + (1-\alpha) \log \alpha - \log \left(\frac{RT}{nFv} \right) - \frac{\alpha(1-\alpha)nF\Delta E}{2.3RT} \quad (4)$$

The k_s value was calculated to be approximately 0.959 s^{-1} (for $v = 1000 \text{ mV s}^{-1}$). This large value signals a high ability of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ for promoting the electrons to the electrode surface.

The electroactive species' the surface coverage (Γ) immobilized on electrode surface can be arrived at from the slope of I_{pc} vs. scan rate ($v < 100 \text{ mV s}^{-1}$) by the following equation [11]:

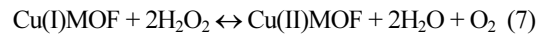
$$\Gamma = (4i_{pc}RT) / (n^2F^2vA) \quad (5)$$

In which i_{pc} is the cathodic peak current (A), n is the number of electrons transferred ($n = 1$ in this case), v is the scan rate (Vs^{-1}), A is the geometric area of the electrode (0.0314 cm^2), and other symbols have their usual meanings. The value of Γ was calculated to be approximately $5.41 \times 10^{-8} \text{ mol cm}^{-2}$ for $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ electrodeposited on a glassy carbon electrode.

3.3. Cyclic Voltammetric Electrochemical Response of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ -GCE Towards H_2O_2

In order to investigate sensing application of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ in hydrogen peroxide detection, an enzyme-free sensor was designed by electrodeposition of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ on a glassy carbon electrode surface. The electrocatalytic activity of $\text{Cu}(\text{btec})_{0.5}\text{DMF}/\text{GCE}$ towards the reduction of H_2O_2 was examined by CV in 0.1 M PBS (pH 6.5) containing 2 mM H_2O_2 . As it can be seen in Fig. 6A, CV curves arrived in 0.1 M PBS with and without H_2O_2 showed the similar current response for the bare GCE

mentioning that the H_2O_2 reduction can rarely be achieved on the GCE surface. In contrast, the cathodic peak current was improved in the presence of hydrogen peroxide at the surface of $\text{Cu}(\text{btec})_{0.5}\text{DMF}/\text{GCE}$, whereas the oxidation peak current decreased. The dramatic increase in reduction current could be attributed to the porosity with high-surface area of $\text{Cu}(\text{btec})_{0.5}\text{DMF}$ film as well as its electrocatalytic behavior to raise the rate of the electron transfer process. With the successive addition of hydrogen peroxide, while anodic peak current has virtually vanished, the cathodic peak current of the modified electrode rose, reflecting a typical electrocatalytic reduction process (EC') (Fig. 6A). The catalytic reduction reaction mechanism can be expressed as follows [12]:



The catalytic currents rose linearly with H_2O_2 con-

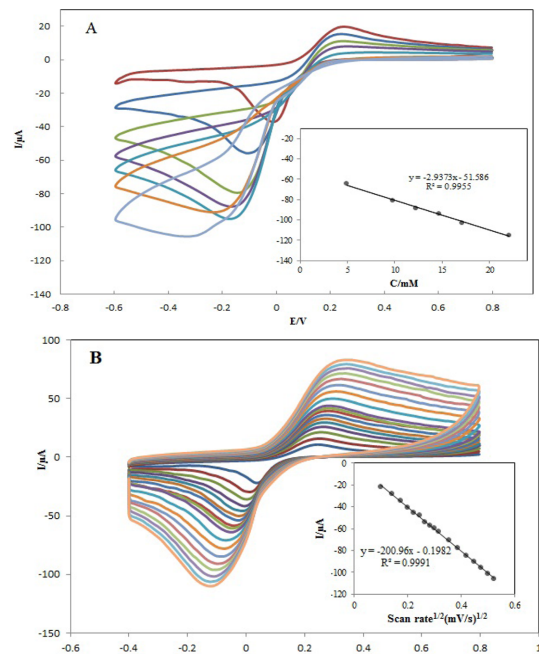


Fig. 6. (A) Cyclic voltammograms of modified glassy carbon in 0.1 M PBS (pH 6.5) containing different concentration of H_2O_2 , (B) cyclic voltammograms in different scan rates in 0.1 M PBS (pH 6.5) containing 2 mM H_2O_2 .

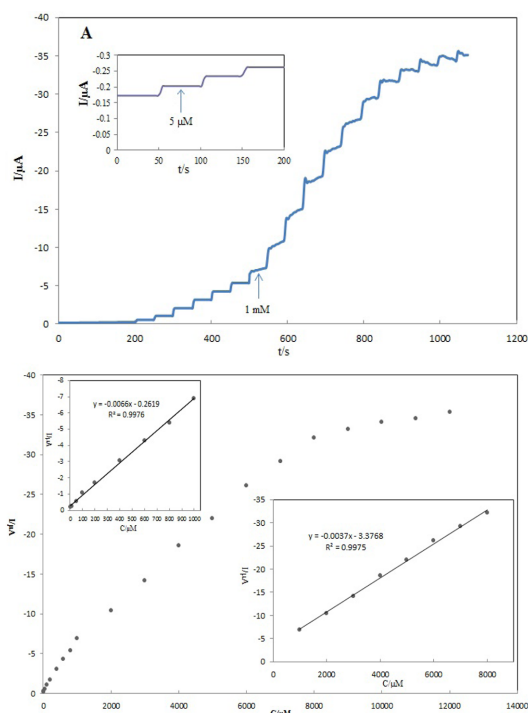


Fig. 7. (A) amperometric response of Cu(btec)_{0.5}DMF-GCE with successive addition of H₂O₂ into 0.1 M PBS, (B) relationship between the reduction current and H₂O₂ concentration.

centration in the range of 5.0–22.0 mM, and the linear regression equation $I_p(\mu\text{A}) = -2.94 [\text{H}_2\text{O}_2](\text{mM}) - 51.59$ with the correlation coefficient (R^2) of 0.995 was obtained. In order to investigate the electrocatalytic mechanism of H₂O₂ reduction at the Cu(btec)_{0.5}DMF/GCE, cyclic voltammograms in 2 mM H₂O₂ were recorded at different scan rates (Fig. 6B). It was concluded that the peak current for the cathodic reduction of H₂O₂ was proportional to the square root of the scan rate, proposing that the process is controlled by diffusion of analyte as can be expected for a catalytic system.

3.4. Chronoamperometric Response of Cu(btec)_{0.5}DMF-GCE to H₂O₂

In order to assess the sensing ability as a non-enzymatic sensor towards the detection of H₂O₂, with a more focus, the typical amperometric responses of the Cu(btec)_{0.5}DMF-GCE was illustrated upon the successive injection of H₂O₂. The amperometric response of Cu(btec)_{0.5}DMF-GCE, with successive

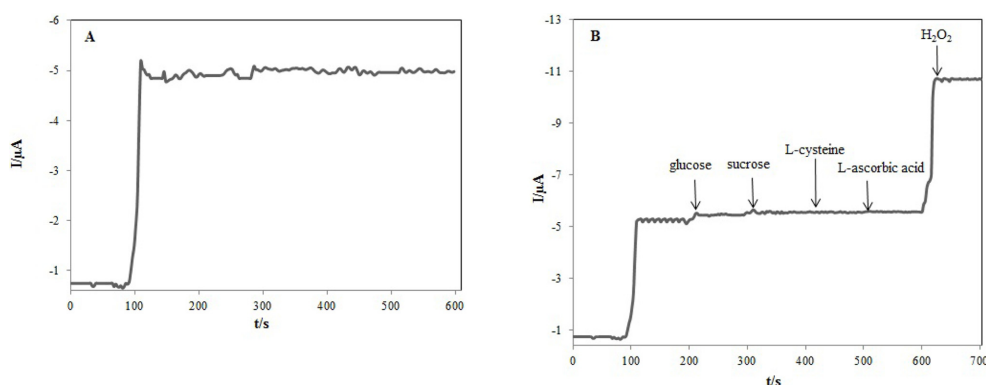
addition of H₂O₂ into 0.1 M PBS (pH 6.5), under the applied potential of -0.2 V in the stirring condition, is shown in Fig. 7. Upon the steeply increasing concentration of H₂O₂, as can be seen in Fig. 7, the response of Cu(btec)_{0.5}DMF-GCE rapidly reaches 95% of the steady-state value within 8 s, which signalled a rapid response and efficient catalytic properties of Cu(btec)_{0.5}DMF-GCE. The Cu(btec)_{0.5}DMF-GCE also showed a good linear relationship between the reduction current and H₂O₂ concentration in the range of 5–1000 μM ($I_p(\mu\text{A}) = -0.007 [\text{H}_2\text{O}_2](\mu\text{M}) - 0.26$ and $R^2 = 0.998$) and 1000–8000 μM ($I_p(\mu\text{A}) = -3.70 [\text{H}_2\text{O}_2](\mu\text{M}) - 3.38$ and $R^2 = 0.998$) (Fig. 7). The values of R^2 show that the regression lines are in line with the experimental data and the regression equations can be applied in the unknown sample determination. The detection limit was calculated according to 3 s/m. For $n = 8$ measurements of background current, the s was obtained 0.0019. By considering of slope of calibration curve ($m = 0.0066$), the detection limit was 0.865 μM which is lower than that in the literatures reported before (Table 1). The detection limit and linear calibration range of the proposed modified electrode were compared with those previously reported, and the results were summarized in Table 1. As can be seen, the analytical parameters are equal or even better than those reported for H₂O₂ detection at the surface of other modified electrodes. It makes clear the excellent performance of the Cu(btec)_{0.5}DMF-GCE as a promising electrochemical sensor in H₂O₂ determination.

3.5. Repeatability, reproducibility, stability and selectivity the H₂O₂ sensor

Repeatability and reproducibility are two important parameters to evaluate the ability of a sensor. A good repeatability is shown by the Cu(btec)_{0.5}DMF-GCE, with a relative standard deviation (RSD) of 7.9% for seven successive detections at 1.0 mM H₂O₂ into 0.1 M PBS (pH 6.5). By the same token, it exhibits a reproducible current response with a RSD of 4.3% for five individual electrodes. These results indicate the excellent reproducibility and repeatability of the Cu(btec)_{0.5}DMF-GC electrode, which are considered a highly desirable characters in its usage in electrochemical non-enzymatic sensors. Furthermore, by measuring the amperometric response of 2 mM H₂O₂, during a prolonged 500 s experiment (Fig. 8A), the stability of the H₂O₂ sensor was tested. The response

Table 1. Comparison of the electrochemical performance of the non-enzymatic H₂O₂ sensors.

Modified electrode	Electrolyte	Analytical method	applied potential (V vs. Ag/AgCl)	Linear range (μM)	LOD (μM)	Ref
Co(pbda)(4,4-bpy)(2H ₂ O) _n /GCE	0.1 M NaOH	Amperometric	-0.40	50-9000	3.76	[13]
Ni(II)-MOF/CNTs	0.1 M NaOH	Amperometric	0.5 (vs Hg/HgO/OH ⁻)	10-51600	2.10	[6]
Cu-MOF/MPC/GCE	0.1 molL ⁻¹ PBS (PH 7.0)	Amperometric	-0.22	10-11600	3.20	[14]
Cu _x ONPs@ZIF-8	0.10 M NaOH	Amperometric	0.7 (vs SCE)	1.5-21442	0.15	[15]
Cu(adp)(BIB)(H ₂ O) _n /GCE	0.1 molL ⁻¹ NaOH	Cyclic voltammetry	-	0.1-2.75	0.07	[16]
Cu-bipy-BTC/MWCNTs/GCE	0.1 molL ⁻¹ PBS (PH 6.0)	Linear sweep voltammety	-	3-70 70-30000	0.46	[17]
CS-SGO@HKUST 1/GCE	25 mM PBS (pH 7.0)	Amperometric	-0.40	1.0-860 860-5600	0.49	[18]
MnO ₂ -ERGO paper	0.1 M PBS (pH 7.4)	Amperometric	-0.5 (vs SCE)	0.1-45.4 mM	10.00	[19]
PdCu/SPCE	0.1 M PBS (pH 7.4)	Amperometric	-0.3	0.5-11 mM	0.70	[20]
Cu(btec) _{0.5} DMF/GCE	0.1 M PBS (pH 6.5)	Amperometric	-0.2	5-1000 μM 1000-8000 μM	0.86	This work

**Fig. 8.** (A) Amperometric response of 2mM H₂O₂ (B) amperometric response of the Cu(btec)_{0.5}DMF-GCE with continuous additions of different compounds.

remains stable throughout the experiment (only a 3.86% decrease in current was observed), showing that Cu(btec)_{0.5}DMF is effectively stabilized onto GC electrode. Finally, the effect of electroactive interfering species on the modified electrode was investigated. Fig. 8B shows the amperometric response of the electrode with the continuous additions of 2 mM H₂O₂, 2 mM glucose, 2 mM sucrose, 2 mM L-cysteine, 2 mM L-ascorbic acid and 2 mM H₂O₂ into 0.1 M PBS. It was found that, the current response of the

mentioned electroactive interfering species is quite negligible which puts a true mark on the fact that the Cu(btec)_{0.5}DMF-GCE possesses a good selectivity for the determination of H₂O₂.

4. Conclusions

In summary, this paper reports on the easy preparation of Cu(btec)_{0.5}DMF to construct a novel non-enzymatic amperometric sensor for the detection of

H₂O₂ for the first time. Electrochemical assays showed that the Cu(btec)_{0.5}DMF modified GC electrode exhibited outstanding performance for non-enzymatic H₂O₂ detection with a wide linear range (5-8000 μM), and a low detection limit (0.865 μM). Moreover, Cu(btec)_{0.5}DMF modified GC electrode demonstrated long-term stability, good reproducibility, and excellent anti-interference ability against common interfering species.

References

- [1] E. Zhou, Y. Zhang, Y. Li and X. He, *Electroanal.*, **2014**, *26(11)*, 2526-2533.
- [2] J. Yang, H. Ye, F. Zhao and B. Zeng, *ACS Appl. Mater. Interfaces*, **2016**, *8(31)* 20407-20414.
- [3] Q. Wang, Y. Yang, F. Gao, J. Ni, Y. Zhang and Z. Lin, *ACS Appl. Mater. Interfaces*, **2016**, *8(47)*, 32477-32487.
- [4] Y. Wang, Y. Wu, J. Xie and X. Hu, *Sens. Actuators B Chem.*, **2013**, *177*, 1161-1166.
- [5] Y. Wang, H. Ge, Y. Wu, G. Ye, H. Chen and X. Hu, *Talanta*, **2014**, *129*, 100-105.
- [6] M.Q. Wang, Y. Zhang, S.J. Bao, Y.N. Yu and C. Ye, *Electrochim. Acta*, **2016**, *190*, 365-370.
- [7] M. Naseri, L. Fotouhi and A. Ehsani, *J. Colloid Interface Sci.*, **2016**, *484*, 314-319.
- [8] P.K. Vabbina, A. Kaushik, N. Pokhrel, S. Bhansali and N. Pala, *Biosens. Bioelectron.*, **2015**, *63*, 124-130.
- [9] R.S. Kumar, S.S. Kumar and M.A. Kulandainathan, *Electrochem. Commun.*, **2012**, *25*, 70-73.
- [10] E. Lavion, *J. Electroanal. Chem.*, **1979**, *101*, 19.
- [11] D.M. Fernandes, C.M.A. Brett and A.M.V. Cavaleiro, *J. Solid State Electrochem.*, **2011**, *15(4)*, 811-819.
- [12] D. Zhang, J. Zhang, R. Zhang, H. Shi, Y. Guo, X. Guo, S. Li and B. Yuan, *Talanta*, **2015**, *144*, 1176-1181.
- [13] L. Yang, C. Xu, W. Ye and W. Liu, *Sens. Actuators B*, **2015**, *215*, 489-496.
- [14] Y. Zhang, X. Bo, C. Luhana, H. Wang, M. Li and L. Guo, *Chem. Commun.*, **2013**, *49(61)*, 6885-6887.
- [15] J. Yang, H. Ye, F. Zhao and B. Zeng, *ACS Appl. Mater. Interfaces*, **2016**, *8(31)*, 20407-20414.
- [16] C. Zhang, M. Wang, L. Liu and X. Yang, *Electrochem. Commun.*, **2013**, *33*, 131-134.
- [17] E. Zhou, Y. Zhang, Y. Li and X. He, *Electroanalysis*, **2014**, *26*, 2526-2533.
- [18] Q. Wang, Y. Yang, F. Gao, J. Ni, Y. Zhang and Z. Lin, *ACS Appl. Mater. Interfaces*, **2016**, *8(47)*, 32477-32487.
- [19] S. Dong, J. Xi, Y. Wu, H. Liu, C. Fu, H. Liu and F. Xiao, *Anal. Chim. Acta*, **2015**, *853*, 200-206.
- [20] A. Uzunoglu, A.D. Scherbarth and L.A. Stanciu, *Sens. Actuators B*, **2015**, *220*, 968-976.